

## REFERENCES

1. M. SILVERMAN and E. A. EVANS, JR., *J. biol. Chem.* **150**, 265 (1943).
2. F. H. JOHNSON and I. LEWIN, *Science, N.Y.* **101**, 281 (1945).
3. F. KRADOLFER and L. NEIPP, *Antibiotics Chemother.* **8**, 297 (1958).
4. L. B. ROBINSON, T. M. BROWN and R. H. WICHELHAUSEN, *Antibiotics Chemother.* **9**, 111 (1959).
5. S. BOLEK, M. SMĚKAL, M. VYKYDAL and Z. ŽIŽKA, *Bratisl. lék. Listy* **45**, 499 (1965).
6. J. CIÁK and F. E. HAHN, *Science, N.Y.* **151**, 347 (1966).
7. R. L. O'BRIEN, J. G. OLENICK and F. E. HAHN, *Proc. natn. Acad. Sci. U.S.A.* **55**, 1511 (1966).
8. A. MILLER and H. C. SMITH, *Br. J. Haemat.* **19**, 417 (1970).
9. G. J. HINE, B. A. BURROWS, L. APT, M. POLLYCOVE, J. F. ROSS and L. A. SARKES, *Nucleonics* **13**, 23 (1955).
10. M. STEINBUCH and M. QUENTIN, *Archs int. Pharmacodyn. Thér.* **110**, 10 (1957).

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**Fluorometric determination of *N*-terminal tryptophan-peptides after formaldehyde condensation**

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TRYPTAMINE can be determined fluorometrically after condensation with formaldehyde followed by oxidation with hydrogen peroxide.<sup>1</sup> The method is non-specific in that also *N*-methyltryptamine and tryptophan give formaldehyde-induced fluorophores with spectral properties indistinguishable from those of the tryptamine fluorophore. Tryptamine, *N*-methyltryptamine and tryptophan are known to react with formaldehyde also on silica gel thin layer.<sup>2-5</sup> It was recently reported that formaldehyde is a sensitive chromatographic detection reagent also for tryptophanyl-dipeptides.<sup>6</sup> From this observation it appeared probable that *N*-terminal-tryptophan-peptides could be determined fluorometrically by a method similar to that described for tryptamine. This assumption was confirmed in the present study.

**Experimental**

Aqueous solutions of tryptamine, tryptophan and various tryptophan-containing dipeptides (see Table 1) were prepared: concentrations of 1-3  $\mu$ g (free base) per ml. Aliquots of these solutions (usually 0.5 ml) were mixed with 0.1 ml formaldehyde solution (18%) in a total volume of 3 ml, made up with 0.1 N sulphuric acid. After heating the samples in boiling water for 20 min, 0.1 ml 5% hydrogen peroxide was added and heating was continued for another 20 min. The samples were cooled in a refrigerator and then analyzed in an Aminco-Bowman spectrophotofluorometer, equipped with an  $x$ - $y$  recorder.

**Results and comments**

All the peptides with an *N*-terminal tryptophan residue gave strong fluorescence after combined treatment with formaldehyde and hydrogen peroxide; no fluorescence resulted if the hydrogen peroxide was omitted. The other tryptophan-containing dipeptides (having the tryptophan amino group engaged in peptide linkage) did not give any fluorescence with the formaldehyde-hydrogen peroxide treatment (Table 1). The spectral properties of the fluorophores of the tryptophanyl-dipeptides were without exception very similar to those of the tryptamine and tryptophan fluorophores (Fig. 1 and Table 1). For tryptophanyl-glycine the fluorescence intensity was proportional to the concentration in the range 0.1 to 3  $\mu$ g/ml (Fig. 2).

The reaction of tryptamine (and tryptophan) with formaldehyde is believed to result in the formation of 1,2,3,4-tetrahydro-norharman which is subsequently dehydrogenated (by the addition of hydrogen peroxide) to 3,4-dihydro-norharman and/or norharman.<sup>1</sup> It appears likely that the same

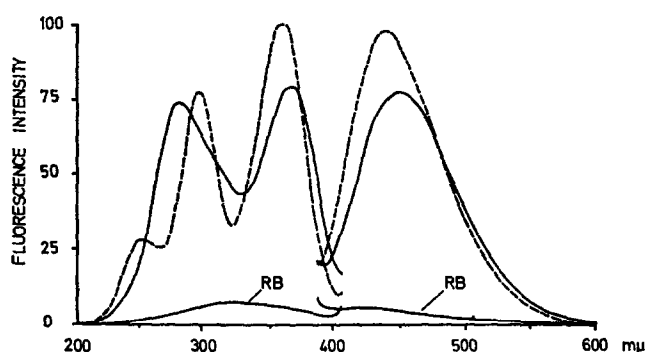


FIG. 1. Excitation (left) and emission (right) spectra of the fluorophores of tryptamine (---) and L-tryptophanyl-L-glycine (—). Equimolar concentration:  $10^{-6}$ M. Reagent blank: RB. Spectra are uncorrected and reproduced as recorded.

TABLE 1. FORMALDEHYDE-INDUCED FLUORESCENCE OF TRYPTAMINE, TRYPTOPHAN AND TRYPTOPHAN-CONTAINING DIPEPTIDES\*

Compounds	Fluorescence intensity of equimolar conc.†	Excitation/emission max. (mμ)‡
Tryptamine	100	(290) 350/440
L-Tryptophan	100	(290) 355/440
L-Tryptophanyl-L-alanine	70	(270) 360/450
L-Tryptophanyl-L-glycine	75	(270) 360/450
L-Tryptophanyl-L-tyrosine	40	(290) 370/460
L-Tryptophanyl-L-tryptophan	30	(290) 370/450
L-Glycyl-L-tryptophan	No fluorescence	
L-Phenylalanyl-L-tryptophan	No fluorescence	
L-Propyl-L-tryptophan	No fluorescence	

\* All peptides were purchased from Miles Laboratories.

† The fluorescence intensity of tryptamine was given as 100.

‡ Uncorrected instrumental values. Minor peak in parenthesis.

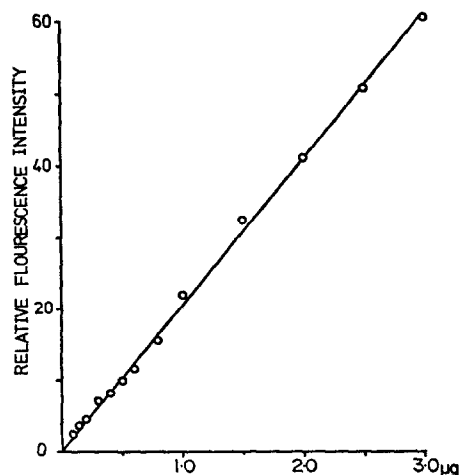
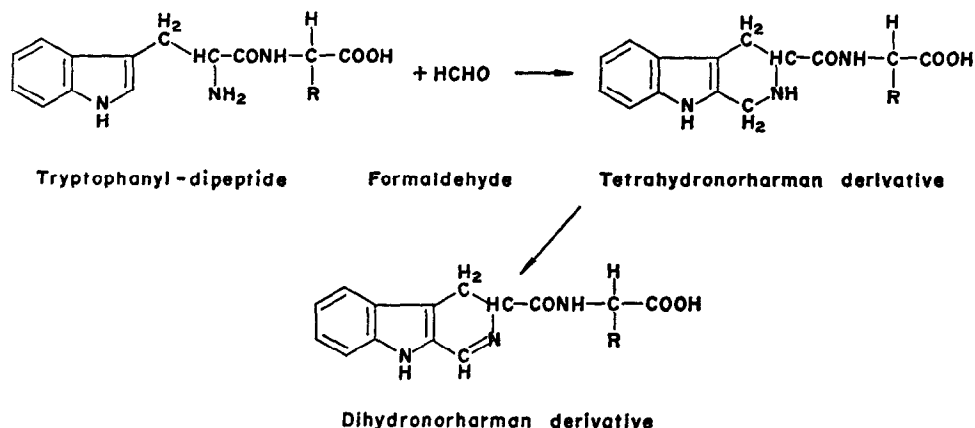


FIG. 2. Rectilinear relationship between the fluorescence intensity and the amount of L-tryptophanyl-L-glycine (in a final volume of 3 ml).

sequence of reactions takes place with the tryptophanyl-peptides according to the following reaction scheme:



Conceivably, all peptides and proteins having an *N*-terminal tryptophan residue will form highly fluorescent conjugates with formaldehyde under the reaction conditions specified. The method has been successfully adapted both for the chromatographic<sup>6</sup> and fluorescence histochemical detection<sup>7</sup> of such peptides.

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#### REFERENCES

1. S. HESS and S. UDENFRIEND, *J. Pharmac. exp. Ther.* **127**, 175 (1959).
2. E. STAHL and H. KALDEWEY, *Z. Physiol. Chem.* **323**, 182 (1961).
3. D. AURES, R. FLEMING and R. HÅKANSON, *J. Chromatog.* **33**, 480 (1968).
4. A. BJÖRKLUND, B. FALCK and R. HÅKANSON, *J. Chromatog.* **47**, 530 (1970).
5. A. BJÖRKLUND, B. FALCK and R. HÅKANSON, *Anal. Biochem.* **35**, 264 (1970).
6. R. HÅKANSON and F. SUNDLER, *J. Chromatog.* in press.
7. R. HÅKANSON and F. SUNDLER, *J. Histochem. Cytochem.* in press.

#### Changes in histaminase content following experimental thermal injury

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HISTAMINASE catalyses the oxidative deamination of histamine and occurs widely in many tissues. Plasma histaminase levels are remarkably increased in pregnancy.<sup>1,2</sup> Histaminase levels have been found, in this laboratory and others, to be raised in myocardial infarction,<sup>3</sup> bronchial asthma<sup>4,5</sup> and anaphylaxis.<sup>6–9</sup> Furthermore, histamine is known to be involved in many tissue challenge situations